

BACTERIOPHAGE GENOMICS: A TOOL FOR LEARNING

BIOINFORMATICS

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Background

Bacteriophages are viruses that infect bacteria. Image 1 depicts a phage's morphology. They are a potential alternative to antibiotics in fighting infections, and their small genome lends themselves well to studying bioinformatics [1]. The Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) program provides undergraduates an opportunity for research-based coursework involving bacteriophages [2]. The students isolate a phage and annotate its genome. The purpose of this project is to outline the process of phage genome annotation; gene 24 from "Damp," a phage isolated by the University of Pittsburgh, depicts the process. Damp is a *Gordonia* phage (i.e., it infects *Gordonia* bacteria). Genome annotation can serve as an introduction to bioinformatics. Bioinformatics involves storing and interpreting biological data (e.g., nucleotide sequences), which can be used to make functional predictions.

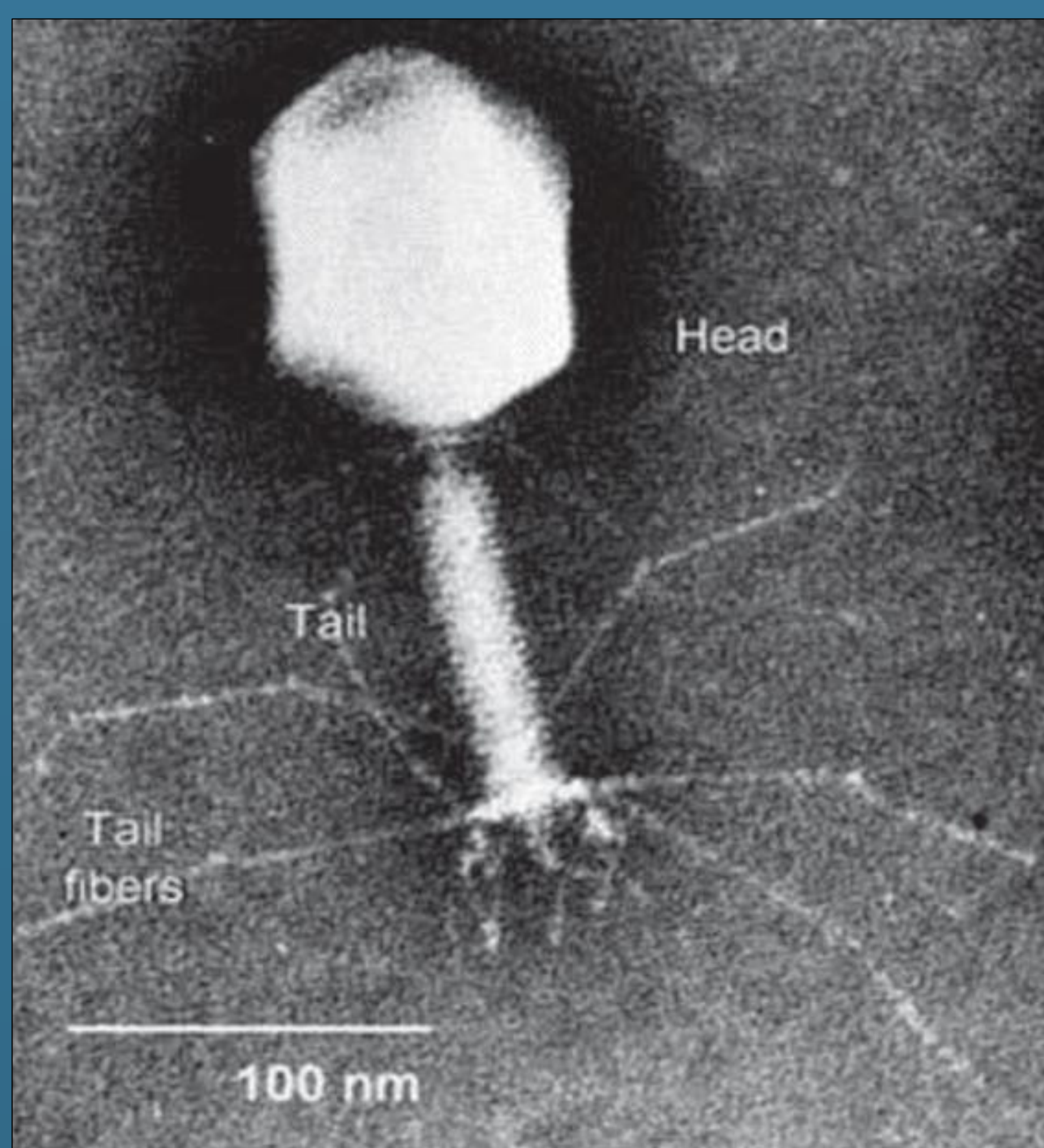


Image 1 – TEM of a bacteriophage. The head contains the DNA, and the tail fibers help bind to the bacteria. Photo: [3]

Manual Annotation

Auto-annotation programs will misinterpret or fail to call 5-10 genes per bacteriophage genome [4]. Because of this, it is paramount that a review of the auto-annotation (i.e., the manual annotation) occurs.

Part 1: Gene Start Prediction

The manual annotation analyzes outputs from various algorithms to confirm or edit the auto-annotation's chosen gene start. Coding potential, gaps, and ribosomal binding site (RBS) scores are assessed.

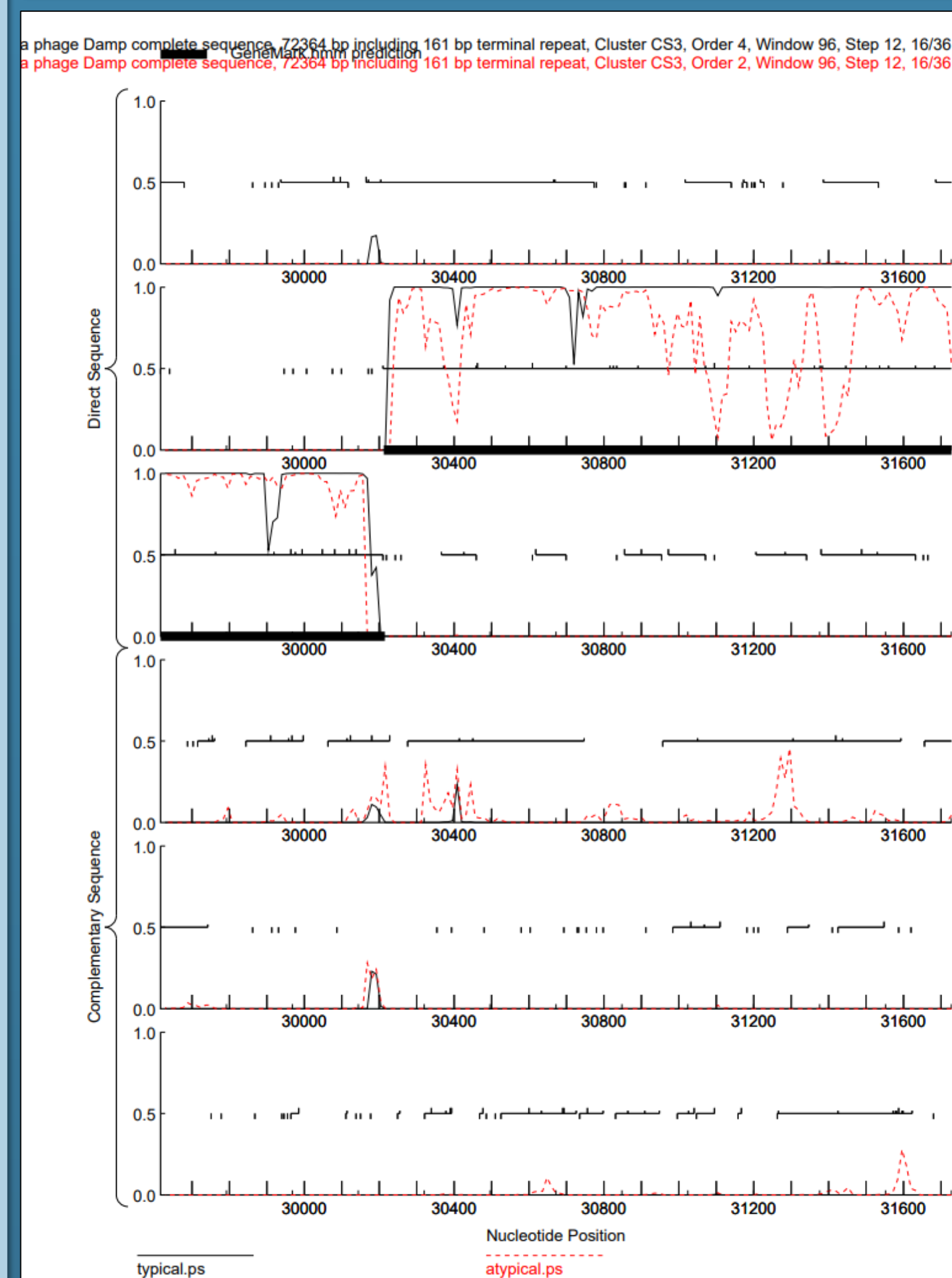


Image 2 (left) – GeneMarkS [5] graph of gene 24 for Damp (Damp_24). The y-axis represents coding potential, the probability that a gene lies at a locus. The x-axis represents the nucleotide position. Gene starts that capture all the coding potential are likely to be the best start for a gene. The graph displays 6 reading frames. A reading frame is the pattern of codons (i.e., triplets of nucleotides) used during translation. The top 3 reading frames are in the forward direction, and the bottom 3 are in the reverse. Red dashed lines represent predictions with an atypical model, and solid lines represent predictions using a typical model; coding potential with the typical model is more robust evidence.

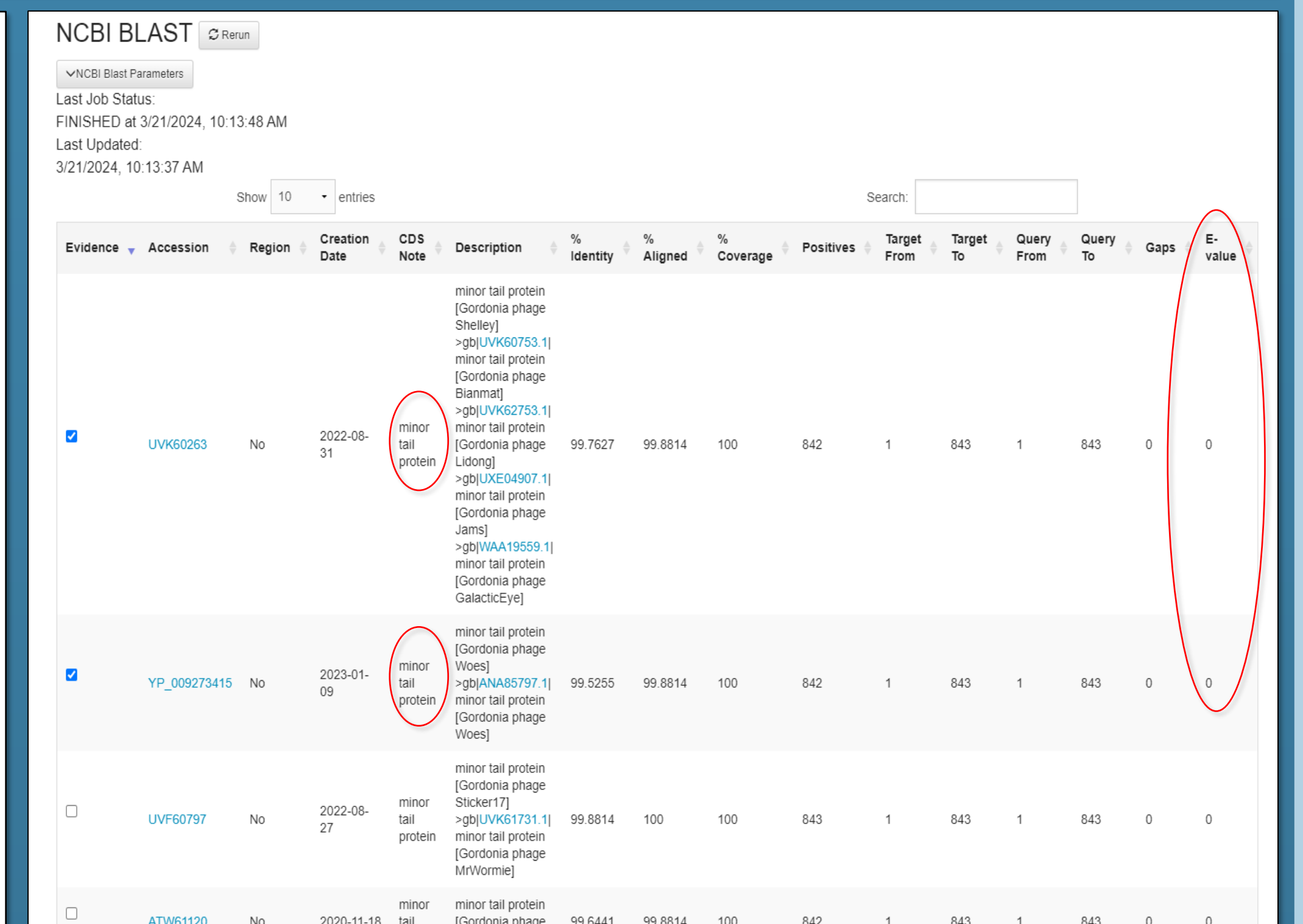
Image 3 (right) – Table of gene starts and their gaps and RBS scores (final score column) [6]. A low gap is stronger evidence for a gene start, as phages favor a compact genome. An RBS score relays the strength of a ribosomal binding site for a gene start. Ribosomal binding sites are sequences of DNA responsible for ensuring translational accuracy. A high score is strong evidence for a gene start.

Direction	Start	Stop	Length	Gap	Spacer	Z-score	Final score	LORF	Start Codon	All GM Coding Capacity	Selected Gene
Forward	30212	32743	2532	-4	9	1.006	-5.492	TRUE	GTG	Yes	<input checked="" type="checkbox"/>
Forward	30374	32743	2370	158	6	2.511	-4.557		GTG		<input type="checkbox"/>
Forward	30461	32743	2283	245	13	1.137	-6.749		GTG		<input type="checkbox"/>
Forward	30464	32743	2280	248	9	1.306	-5.935		ATG		<input type="checkbox"/>
Forward	30539	32743	2205	323	8	1.507	-6.041		GTG		<input type="checkbox"/>
Forward	30611	32743	2133	395	13	0.93	-7.183		ATG		<input type="checkbox"/>
Forward	30665	32743	2079	449	8	1.94	-5.235		TTG		<input type="checkbox"/>
Forward	30698	32743	2046	482	16	1.879	-5.937		TTG		<input type="checkbox"/>
Forward	30701	32743	2043	485	14	1.012	-7.314		GTG		<input type="checkbox"/>
Forward	30818	32743	1926	602	7	1.726	-5.996		GTG		<input type="checkbox"/>

Part 2: Protein Function Prediction

After predicting where a gene starts, the gene product (i.e., the protein) can be assessed. This involves matching nucleotide or amino acid sequences against databases.

Image 4 (right) – The NCBI basic local alignment search tool (BLAST) [7] matches a gene's nucleotide sequence against the NCBI database. It may target an entire gene or a portion of one. Matches with known gene products indicate possible protein function. The e-value for each hit represents the probability that an alignment occurred by chance. Therefore, hits with lower e-values are stronger evidence for function.



Nr	Hit	Name	Probability	E-value	Score	SS	cols	Length
1	80DZ_D	Interleukin-12 receptor subunit beta-2, Calmodulin-1; Complex, Cytokine, Receptor, SIGNALING PROTEIN; HET: BMA, NAG, MAN;	99.75	5.6e-14	168.57	44.9	501	769
2	8G4L_am	Titin; cardiac, myosin, filament, complex, CONTRACTILE PROTEIN;(Homo sapiens)	99.68	2.2e-12	161.64	47.6	316	1084
3	4YH7_A	Receptor-type tyrosine-protein phosphatase delta; Trans-synaptic complex, Synapse organizer, HYDROLASE-IMMUNE SYSTEM com	99.66	5e-12	154.12	46.4	386	841

Image 5 – HHpred [8] matches amino acid sequences against protein databanks. It uses a different algorithm than NCBI BLAST. HHpred is important to protein calls, as it may align to wet lab results, which is strong evidence. HHpred also provides the e-value for a particular hit. This query for Damp_24 is matching to a contractile protein, which is evidence for a minor tail functional call.

References

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